

Analysis of *Bacteroides fragilis* Clinical Strains Isolated in Kazakhstan

Microbiology[®]

Resource Announcements

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ABSTRACT Our aim was to study the nucleotide sequences of 9 previously undescribed strains of *B. fragilis* collected from patients with intra-abdominal diseases at city hospitals in Nur-Sultan, Kazakhstan.

B acteroides fragilis is a commensal bacterium that is found in the intestines of most people and that can become an opportunistic pathogen. Amounting to only about 0.1 to 0.5% of the total bacterial mass of the intestine, *B. fragilis* is the most frequently isolated anaerobe from clinical samples obtained from deep intra-abdominal abscesses, purulent skin infections, soft tissue infections (1, 2), diarrhea, and colorectal cancer (3).

B. fragilis strain no. 4 to 12 were isolated from clinical samples of patients diagnosed with acute peritonitis receiving treatment at City Hospitals No. 1 and No. 2 and Regional General Hospital No. 2. Informed consent and questionnaires were approved by the local ethics committee of the RSE National Center for Biotechnology of the Ministry of Education and Science of the Republic of Kazakhstan (extract from protocol no. 4 of 29 August 2017).

Samples were collected from the drainage wounds by use of swabs with subsequent immersion of probes into tubes containing Amies medium. After that, samples were cultivated on Bacteroides bile esculin agar (BBE; Conda) at 37°C for 72 h under anaerobic conditions (an aerostat, gas pack). Isolates were identified using matrix-assisted laser desorption ionization-time of flight mass spectrometry (MALDI-TOF MS; Bruker Daltonics, Bremen, Germany). Bacterial colonies growing on solid medium were removed with a sterile plastic tip and resuspended in 1,000 μ l of Tris-EDTA (TE) buffer. Total DNA was extracted from all strains using the cetyltrimethylammonium bromide (CTAB) method (4). Then, 150 ng of the total genomic DNA from each isolate of B. fragilis was used for sequencing. Library preparation and Illumina MiSeq sequencing were performed using the Nextera DNA Flex library prep kit and a MiSeg reagent kit v3 with 300-bp paired-end reads (600 cycles) according to the manufacturer's instructions. Quality assessment of the sequencing data (in FASTQ format) was done using FastQC v0.11.15 (5), followed by trimming of adapters and low-quality bases with a Phred quality score of less than 20 using Trimmomatic (6). Genomes were assembled using the SPAdes assembler v3.13.2 using a k-mer length of 127 with the "--careful" mode (7). Comparative phylogenetic analysis was performed with the CSI Phylogeny v1.4 tool from whole-genome sequences using the following parameters: minimum depth, 80×; minimum relative depth, 100%; minimum distance between single-nucleotide polymorphisms (SNPs), 1,000 bp; minimum SNP quality, 500; minimum read mapping

Citation Zholdybayeva E, Kozhahmetova S, Tarlykov P, Atavliyeva S, Mukhtarova K, Syzdykov T, Khasenov R, Shevtsov A, Amirgazin A, Daniyarov A, Ramankulov Y. 2021. Analysis of *Bacteroides fragilis* clinical strains isolated in Kazakhstan. Microbiol Resour Announc 10:e01311-20. https://doi.org/10 .1128/MRA.01311-20.

Editor David Rasko, University of Maryland School of Medicine

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Received 17 November 2020 Accepted 9 January 2021 Published 4 February 2021



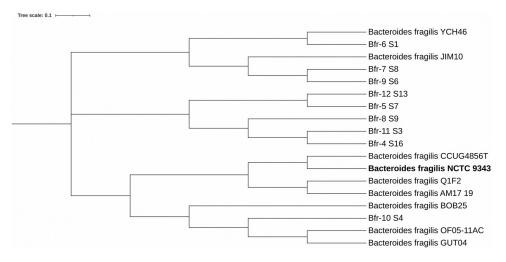


FIG 1 Phylogram of available *Bacteroides fragilis* genomes based on the concatenated alignment of the highquality SNPs using CSI Phylogeny v1.4. The following available genomes were retrieved from GenBank (ncbi .nlm.nih.gov/genome/): *Bacteroides fragilis* strain JIM10 (GenBank accession no. CM004507.1), *Bacteroides fragilis* strain YCH46 (NC_006347.1), *Bacteroides fragilis* strain CCUG4856T (CP036555), *Bacteroides fragilis* strain Q1F2 (NZ_CP018937.1), *Bacteroides fragilis* strain AM17-19 (NZ_QRJX0000000.1), *Bacteroides fragilis* strain BOB25 (CP011073.1), *Bacteroides fragilis* strain OF05-11AC (NZ_QSWE00000000.1), and *Bacteroides fragilis* strain GUT04 (CP043610.1). The reference genome, *Bacteroides fragilis* NCTC 9343 (NC_003228.3), is indicated in bold.

quality, 500; and minimums Z-score, 3.29 (8). We discarded sequences with low quality, i.e., ambiguous bases. Figure 1 was created using iTOL v4 (9). *B. fragilis* strain NCTC 9343 (GenBank accession no. NC_003228.3) was used as a reference sequence. Genome annotation was performed using PGAP v4.11 (10). Default parameters were used for all software.

Data from the whole-genome sequencing of the 9 clinical *B. fragilis* strains are presented in Table 1.

Results of the phylogenetic analysis are shown in Fig. 1. *B. fragilis* Bfr-10 is located farther from all the strains studied.

Data availability. The 9 whole-genome shotgun projects have been deposited at DDBJ/ENA/GenBank under the accession numbers listed in Table 1.

Strain	Strain	Genome	Genome	No. of	Total no.	Avg read		G+C	
no.	name	size (bp)	coverage (×)	contigs	of reads	length (bp)	N ₅₀ (bp)	content (%)	Accession no./SRA no.
4	Bfr-4	4,920,846	20.0	128	1,724,380	192	69, 701	43.41	JACEFH000000000/ SRX9403175
5	Bfr-5	5,219,927	38.565	77	1,555,229	221	181, 129	43.36	JACENG00000000/ SRX9403182
6	Bfr-6	5,259,406	49.394	41	1,398,871	231	378, 644	43.33	JACFSS00000000/ SRX9404153
7	Bfr-7	5,175,039	19.59	28	736,748	273	379, 532	43.25	JACFST00000000/ SRX9404154
8	Bfr-8	5,243,270	64.17	40	1,258,909	242	311, 507	43.24	JACFSU000000000/ SRX9404155
9	Bfr-9	5,125,231	29.757	53	1,501,107	218	290, 748	43.45	JACFSV00000000/ SRX9404156
10	Bfr-10	5,242,965	31.158	36	1,511,363	226	397, 964	43.12	JACFSW000000000/ SRX9404157
11	Bfr-11	5,236,934	33.68	32	1,386,003	245	407, 555	43.4	JACFSX000000000/ SRX9404158
12	Bfr-12	5,321,320	31.143	92	1,272,842	229	182, 180	43.39	JACFSY000000000/ SRX9404159

TABLE 1 Genome characteristics of 9 B. fragilis strains

ACKNOWLEDGMENTS

We thank the I. Azizov Institute of Antimicrobial Chemotherapy, Smolensk State Medical University (Smolensk, Russia), for scientific support.

The study was supported by grant funding of the Committee of Science of the Ministry of Education and Science of the Republic of Kazakhstan within the framework of the project "Study and Assessment of Sensitivity of *Bacteroides fragilis* to Broad-Spectrum Antibiotics in Patients with Intra-abdominal Anaerobic Infections" (AP05132131).

REFERENCES

- Valguarnera E, Wardenburg JB. 2020. Good gone bad: one toxin away from disease for *Bacteroides fragilis*. J Mol Biol 432:765–785. https://doi .org/10.1016/j.jmb.2019.12.003.
- Jamal W, Khodakhast FB, Azmi AA, Sóki J, Hashem GA, Rotimi VO. 2020. Prevalence and antimicrobial susceptibility of enterotoxigenic extra-intestinal *Bacteroides fragilis* among 13-year collection of isolates in Kuwait. BMC Microbiol 20:14. https://doi.org/10.1186/s12866-020-1703-4.
- Keenan JI, Aitchison A, Purcell RV, Greenlees R, Pearson JF, Frizelle FA. 2016. Screening for enterotoxigenic *Bacteroides fragilis* in stool samples. Anaerobe 40:50–53. https://doi.org/10.1016/j.anaerobe.2016.05.004.
- 4. Sambrook J, Russell DW. 2001. Molecular cloning. A laboratory manual. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY.
- Andrews S. 2014. FastQC: a quality control tool for high throughput sequence data. https://www.bioinformatics.babraham.ac.uk/projects/fastqc/.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. https://doi.org/10 .1093/bioinformatics/btu170.

- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.
- Kaas RS, Leekitcharoenphon P, Aarestrup FM, Lund O. 2014. Solving the problem of comparing whole bacterial genomes across different sequencing platforms. PLoS One 9:e104984. https://doi.org/10.1371/ journal.pone.0104984.
- Letunic I, Bork P. 2019. Interactive Tree Of Life (iTOL) v4: recent updates and new developments. Nucleic Acids Res 47:W256–W259. https://doi .org/10.1093/nar/gkz239.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44:6614–6624. https://doi.org/10.1093/nar/gkw569.